

Gavrylenko Yu. V.

Shupyk National Medical Academy of Postgraduate Education, Kyiv, Ukraine

## The impact of BNO 1030 on phagocytic activity of white blood cells in rats with type 1 diabetes mellitus

For cite: Mezhdunarodnyi Endokrinologicheskii Zhurnal. 2017;13:374-9. doi: 10.22141/2224-0721.13.5.2017.110028

**Abstract. Background.** In the pathogenesis of type 1 diabetes mellitus (DM1) and many of its complications, an important place belongs to a violation of the functional capacity of the immune system, which is the subject of numerous studies. Considering the effectiveness of immunomodulating therapy for DM1 and its complications, one of the most important problems is the search for new effective and harmless immunocorrecting agents with high pharmacological activity. The purpose of this study was to evaluate the effect of the phytopreparation BNO 1030 on the phagocytic activity of blood leukocytes in rats with DM1. **Materials and methods.** Experimental DM1 in rats was induced by a single intraperitoneal injection of streptozotocin. The percentage of phagocytic cells and macrophage uptake of live fluorescent bacteria in the samples were determined using fluorescent live bacteria *Escherichia coli*. **Results.** The effect of the drug BNO 1030 on the phagocytic activity of white blood cells with DM1 in rats was investigated. Under hyperglycemia it was found changes in the redistribution of the main types of leukocytes, namely granulocytes and agranulocytes. The administration of the drug BNO 1030 to both control and diabetic animals resulted in an increase in the number of agranulocytes, which may indicate a modulating effect of the drug on the immune system of animals. Under these conditions, phagocytic activity of leukocytes was decreased as phagocytic number in group of diabetic rats was reduced by 42 % in comparison with group of control animals. BNO 1030 administration to diabetic rats caused an increase of phagocyte number by 24 % compared to the group of diabetic animals. These changes, in turn, are also accompanied by a decrease of phagocytic index as in diabetes, and when drug was administered to experimental groups, indicating disorders in nonspecific cellular immune system. **Conclusions.** BNO 1030 due to its immunomodulatory effect is effective in the treatment of DM1 and its comorbidity with other chronic diseases.

**Keywords:** experimental type 1 diabetes mellitus; BNO 1030; white blood cells; phagocytic activity; phagocytic number; phagocytic index

### Introduction

Type 1 diabetes mellitus (DM1) is a multifactorial and chronic endocrine disease characterized by losing of insulin-producing pancreatic  $\beta$ -cells, which occurs as a result of an autoimmune reaction by forming autoantibodies and autoreactive T-lymphocytes in these cells factors [8, 13, 15]. Patients with DM1 have an increased susceptibility to infections, especially viral ones, which is associated with the inhibition of the protective functions of the body as a result of immune system disorders, increased cell adhesion of microorganisms, the presence of micro- and

macroangiopathy, neuropathy, and the development of other pathogenetic processes [7, 12, 16]. In the pathogenesis of DM1 and many of its complications, an important place belongs to the violation of the functional capacity of the immune system, which is the subject of numerous studies [4, 6]. Patients with DM1 have significant changes in chemotaxis, a decrease in the bactericidal activity of neutrophils, increased production of reactive oxygen species, leukotrienes, secretion of lysosomal enzymes, and changes in the level of intracellular calcium are often observed [2, 16].

Diseases of the ENT organs and respiratory tract are frequent reasons for seeking medical help. A special attention and methodological approach requires the treatment of DM1 in the case of its combination with other chronic diseases, for example, with chronic tonsillitis (ChT). ChT increases metabolic and functional disorders in the body, and also leads to decompensation of carbohydrate metabolism and even to ketoacidosis, which deteriorate the pathological process in the tonsils [1, 3]. In conditions when the development and course of DM1 is complicated by the activation of foci of infection and often septic states, the evaluation of the functional activity of phagocytes, in which neutrophils dominate, may allow a more correct justification of the use of efferent therapy [1, 5].

Considering the effectiveness of immunomodulatory therapy for DM1 and its complications, one of the most important problems is the search for new effective and harmless immunocorrecting agents with high pharmacological activity. Therefore, it was relevant to study the effect of BNO 1030 Bionorica AG (Germany) on the functional and phenotypic characteristics of cells of tonsil in patients with chronic tonsillitis *in vitro*. Accordingly, one possible explanation for the abnormal leukocyte function in diabetes mellitus might be a down-regulation of adhesion molecules that regulate leukocyte recruitment during the course of inflammatory processes. The impaired local exudative cellular reaction in alloxan-induced diabetic rats is a consequence of defective leukocyte-endothelial interactions. Intravital microscopic examination of the internal spermatic fascia microcirculatory network showed that a reduced number of leukocytes rolling along the venular endothelium is observed from the early stages of diabetes. Considerable support to these clinical investigations was given by experimental studies on diabetic rats. Insulin restores the appropriate response to injury through a direct or indirect action on endothelial cells and leukocytes. For future studies the challenge remains to better understand the integration of adaptive immune systems with the endocrine system, providing new insights into how inflammation is regulated.

**The purpose of this study** was to evaluate the effect of the BNO 1030 on the phagocytic activity of blood leukocytes in rats with DM1.

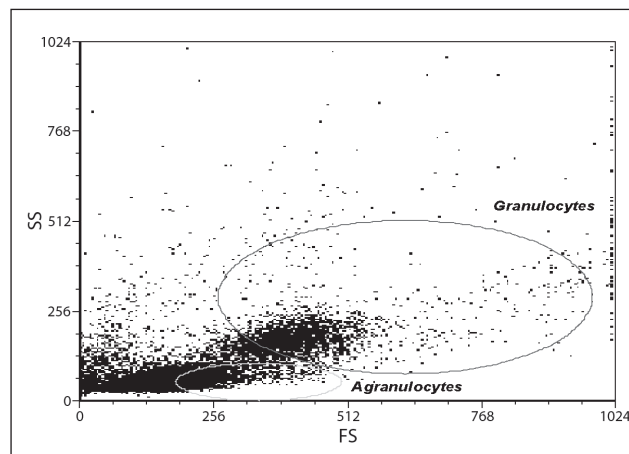
## Materials and methods

The studies were carried out on intact male Wistar rats weighing 130–150 g. The maintenance of animals and conducting experiments with them were carried out in accordance with generally accepted international requirements for work with experimental animals and in accordance with the relevant national provisions for conducting experimental work [4]. The experimental animals were kept on a standard ration of the vivarium, with free access to food and water. Experimental DM1 in rats was induced by a single intraperitoneal injection of streptozotocin (Sigma-Aldrich Co. LLC, USA) with a dose of 55 mg/kg body weight, diluted in 0.1 M

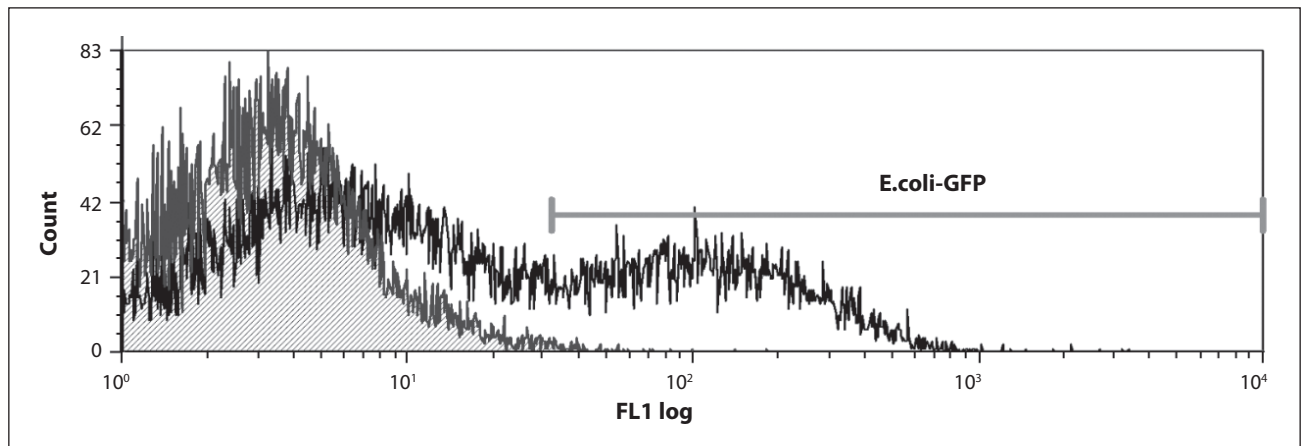
citrate buffer, pH 4.5 before administration to animals. The rats of the control group of the same age were intraperitoneally injected with 0.5 ml 0.1 M citrate buffer, pH 4.5. Blood sampling in animals was performed in the morning after fasting (12 hours) from the retrobulbar venous sinus of the eye, under light ether anesthesia. The blood glucose level was determined using a Precision Xtra Plus blood glucose meter (MediSense UK Ltd., UK). Leukocytes were obtained on the day of the experiment from peripheral blood of experimental animals by osmotic shock of erythrocyte membranes. To this end, the heparinized blood was mixed with a cold lysis solution (0.15 mol/L NH<sub>4</sub>Cl, 1 mmol/L KHCO<sub>3</sub>, 0.1 mmol/L EDTA, pH 7.2–7.4) in a ratio of 1 : 20 and after a thorough shaking incubated for 10 minutes at 37 °C. After the lysis time of the erythrocytes, the samples were centrifuged (400 g, 5 min) for the precipitation of leukocytes. The supernatant was removed and the pellet was washed twice with saline by centrifugation (400 g, 5 min). The washed pellet was resuspended at a concentration of  $2 \times 10^6$  cells/ml in phosphate buffered saline (PBS) (Phosphate buffered saline, pH 7.2). Blood serum was obtained by centrifugation of whole blood in a centrifuge Eppendorf 5810R (USA) at 1300 g for 7 min at 22 °C. Serum was stored at –72 °C until use.

Evaluation of redistribution between different leukocyte populations was carried out using two parameters of the COULTER EPICS XL protocol cytofluorimeter (Beckman Coulter, USA) equipped with an argon laser ( $\lambda = 488$  nm): direct (FS, cell size) and lateral light scattering (SS, granularity Cells), Fig. 1.

The percentage of phagocytic cells and macrophage absorption by live fluorescent bacteria in the samples were determined using fluorescent live bacteria *Escherichia coli*, according to the method of [9]. The cells were incubated for 1 hour at 37 °C with live fluorescent bacteria of *Escherichia coli* at a concentration of  $6 \times 10^6$  bacteria/ml in 1% BSA/PBS buffer. After washing in 1% BSA/PBS buffer, an analysis was made of the percentage of phagocytic cells in the flow cytofluorimeter, Fig. 2.



**Figure 1. A typical histogram of the distribution between leukocyte populations in terms of the direct (FS) and lateral light scattering (SS)**



**Figure 2. Histograms reflecting the fluorescence of macrophage cells with the FL3 channel. The blue color is marked with macrophage cells without the addition of fluorescent bacteria *E.coli***

The principle of the method for determining phagocytic activity is based on the absorption of *E.coli* by phytochemicals, monocytes and macrophages (capable of expressing Green Fluorescent Protein (GFP) with a molecular weight of 26.9 kDa by the phagocytosis). This protein fluoresces with green light when excited by light blue (laser, 488 nm). Accordingly, the more bacteria are absorbed by the cell, the higher the fluorescence intensity of GFP in it.

The statistical processing of the obtained results was carried out using the Statistica 6.0 program. Samples were compared using Student’s t-test. The results are presented as the mean value (M) and the standard error of the mean value ( $\pm m$ ). The difference was considered statistically significant at  $p < 0.05$ .

**Results**

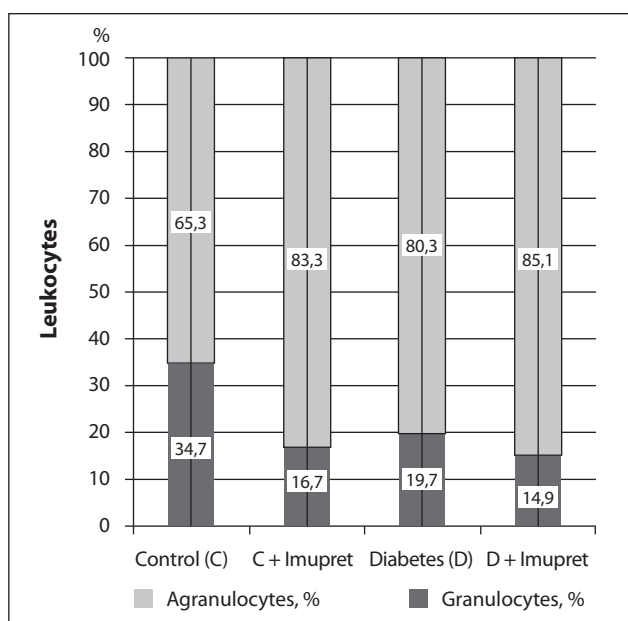
DM1 occurs as a result of autoimmune inflammation and caused by various exogenous degradation factors of  $\beta$ -cells of the pancreas, accompanied by

numerous complications due to relative and/or absolute insufficiency of insulin [15, 19]. A characteristic feature of the development of DM1 is a disruption of the metabolism of carbohydrates, which results in an increase in blood glucose concentration. Therefore, to validate the experimental model of DM1, it was important to determine the level of glucose in the blood of the animals. At the beginning of the experiments, blood glucose levels were almost the same in all the studied groups, but after six weeks of DM-1 development, glucose levels in the blood increased 2.3-fold compared to control animals, which is a confirmation of the development of uncompensated hyperglycemia in animals.

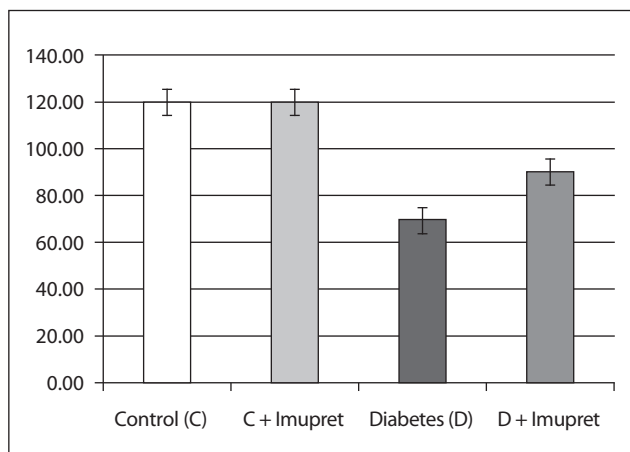
As evidenced by the data obtained, shown in Fig. 3, changes in the redistribution of leukocytes were detected. Thus, the amount of agranulocytes in the blood of diabetic rats increased by 15 % compared with the control animals, while the number of granulocytes, on the contrary, decreased. The revealed changes in their redistribution indicate the reaction of the hematopoietic apparatus to the development of pathological processes in the organism of animals induced by diabetes mellitus, which is consistent with the literature data, which indicate that changes in the distribution of the main types occur at various blood diseases and the appearance of other pathological conditions [10, 14].

When Imupret was administered to both control and diabetic rats, an increase in the amount of agranulocytes was also observed. Since the components of the agglutination are lymphocytes and monocytes, the data obtained may indicate modulating the effect of the drug on the immune system.

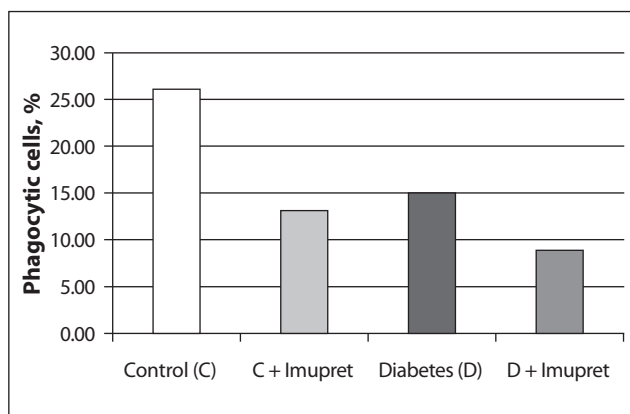
For a more complete evaluation of phagocytic activity, the phagocytic number (the average number of microbes that are absorbed by one phagocyte) was determined. The obtained data indicate that the phagocyte count in the group of rats with DM1 significantly decreases, namely, less than 42 % compared to the control animals, Fig. 4. When administered to control rats of the drug BNO 1030, no reliable changes in the magnitude of the phagocytic number were



**Figure 3. Redistribution of blood leukocytes in groups of experimental animals**



**Figure 4. The phagocytic number is expressed in the relative intensity of the fluorescence of *Escherichia coli* in one cell (n = 5)**



**Figure 5. Phagocytic index of leukocytes (n = 3)**

detected. However, in the group of diabetic rats receiving BNO 1030, an increase in phagocytosis was found to be 24 % compared to a group of diabetic animals that did not receive the drug.

It was observed both in diabetes and in the administration of the drug to experimental animals in comparison with the control group evaluating the phagocytic index (the ratio of phagocytic cells to the total number of cells in leukocytes) (Fig. 5). It should be noted that the observed decrease in the phagocytic index correlates with the total content of granulocytes in the samples (Fig. 3), taking into account the redistribution between the two major types of leukocytes in the blood at the experimental DM1. The decrease of the investigated parameters of activity of phagocytosis testifies to violations in the system of non-specific cellular immunity. This can be the result of reduced production of phagocytes, their rapid decay, violation of their mobility, violation of absorption and neutralization of foreign agents, and the like.

## Discussion

According to the obtained data, it becomes apparent that hyperglycemia leads to a decrease in the activity of the immune system, which agrees with the results of other authors who found that patients with

diabetes form a risk group for susceptibility to infections as a result of reduced immunity [8, 11].

Studies with diabetic rats and also showed a decreased neutrophil migration, phagocytosis capacity and hydrogen peroxide production. Furthermore, the reduction of blood glucose levels by insulin treatment of diabetic patients or rats has been reported to be significantly correlated with improvement of neutrophil phagocytosis capacity. During an inflammatory response, leukocytes roll along the lining endothelium of post-capillary venules and eventually become firmly attached to the vascular wall before migrating into tissues. Specific adhesion glycoproteins expressed on the surface of leukocytes and endothelial cells play a relevant role in the accumulation of leukocytes in the inflammatory lesion. Members of the selectin family of cell adhesion molecules are thought to mediate leukocyte rolling along the walls of the microvasculature.

Another important advance in understanding the pathogenesis of neutrophil dysfunction and inflammatory disorders in diabetes is the observation that glucose or its analogues interact with proteins or lipids. The end products of this non-enzymatically catalyzed reaction, termed advanced glycation (glycosylation or glycoxidation) end products (AGEs), have been linked to the development of long-term complications of diabetes. Early glycation and oxidation processes result in the formation of reversible Schiff bases, which undergo an intramolecular rearrangement to form the Amadori products like glycated hemoglobin (HbA1c) that is elevated in diabetic patients. A small proportion of these products undergo further slow and irreversible chemical rearrangements to form AGEs, which accumulate in the vasculature under conditions that are accelerated during hyperglycemia and when protein turnover is delayed.

## Conclusions

Thus, the data obtained indicate a pronounced reduction in the body's resistance to various infections, especially when they occur on the background of DM1.

A confirmation of this is the process of phagocytosis, which undergoes significant changes in patients with DM1.

These changes, in the light of experimental data, partially prevent the use of the drug BNO 1030, which is able to exhibit immunomodulatory effect in patients with DM1.

1. It was established that in the experimental DM1 changes in the redistribution of the main types of leukocytes occur. When BNO 1030 was administered to control and diabetic animal, the amount of agranulocytes increased, which may indicate a modulating effect of the drug on the immune system.

2. The phagocyte count in the group of DM1 patients was significantly lower and decreased by 42 % compared with the control animals. When administered to diabetic rats, BNO 1030 showed an increase in phagocyte count by 24 % compared to a group of diabetic animals that did not receive the drug.



3. The reduction of the phagocytic index in both groups indicates a violation of the system of nonspecific cellular immunity

**Conflicts of interests.** Author declares the absence of any conflicts of interests that might be construed to influence the results or interpretation of their manuscript.

## References

1. Gavrilenko IuV. Features of upper respiratory tract lesions in children and adolescents with type 1 diabetes mellitus. *Sovremennaya pediatriya*. 2015;7(71):62-5; doi 10.15574/SP.2015.71.62 (in Russian).
2. Laiko AA, Gavrylenko IuV. Pattern of ENT-organ lesions in children with 1 type diabetes mellitus. *Rinologiya*. 2014;1:61-5. (in Ukrainian).
3. Pertseva NO, Martsynik EN, Chursinova TV. Features of insulin resistance in patients with long history of type 1 diabetes mellitus, methods of its correction. *Mezhdunarodnyi Endokrinologicheskii Zhurnal*. 2017;13(1):23-7. doi 10.22141/2224-0721.13.1.2017.96750. (in Russian).
4. Adeghe E, Schattner P, Dunn E. An update on the etiology and epidemiology of diabetes mellitus. *Ann N Y Acad Sci*. 2006 Nov;1084:1-29. doi: 10.1196/annals.1372.029.
5. Bicker H, Höflich C, Wolk K, Vogt K, Volk HD, Sabat R. A simple assay to measure phagocytosis of live bacteria. *Clin Chem*. 2008 May;54(5):911-5. doi: 10.1373/clinchem.2007.101337.
6. Casqueiro J, Casqueiro J, Alves C. Infections in patients with diabetes mellitus: A review of pathogenesis. *Indian J Endocrinol Metab*. 2012 Mar;16 Suppl 1:S27-36. doi: 10.4103/2230-8210.94253.
7. Chattopadhyay S, Ramanathan M, Das J, Bhattacharya SK. Animal models in experimental diabetes mellitus. *Indian J Exp Biol*. 1997 Nov;35(11):1141-5. PMID: 9567740.
8. Bilgic S, Aktas E, Salman F, et al. Intracytoplasmic cytokine levels and neutrophil functions in early clinical stage of type 1 diabetes. *Diabetes Res Clin Pract*. 2008 Jan;79(1):31-6. doi: 10.1016/j.diabres.2007.06.011.
9. Valle A, Giamporcaro GM, Scavini M, et al. Reduction of circulating neutrophils precedes and accompanies type 1 diabetes. *Diabetes*. 2013 Jun;62(6):2072-7. doi: 10.2337/db12-1345.
10. Knip M, Veijola R, Virtanen SM, Hyöty H, Vaarala O, Akerblom HK. Environmental triggers and determinants of type 1 diabetes. *Diabetes*. 2005 Dec;54 Suppl 2:S125-36. PMID: 16306330.
11. James S, Gallagher R, Dunbabin J, Perry L. Prevalence of vascular complications and factors predictive of their development in young adults with type 1 diabetes: systematic literature review. *BMC Res Notes*. 2014 Sep 2;7:593. doi: 10.1186/1756-0500-7-593.
12. Zhang C1, Yang J, Jennings LK. Leukocyte-derived myeloperoxidase amplifies high-glucose--induced endothelial dysfunction through interaction with high-glucose--stimulated, vascular non--leukocyte-derived reactive oxygen species. *Diabetes*. 2004 Nov;53(11):2950-9. PMID: 15504976.
13. Szablewski L, Sulima A. The structural and functional changes of blood cells and molecular components in diabetes mellitus. *Biol Chem*. 2017 Apr 1;398(4):411-423. doi: 10.1515/hsz-2016-0196.
14. Popov D. Endothelial cell dysfunction in hyperglycemia: Phenotypic change, intracellular signaling modification, ultrastructural alteration, and potential clinical outcomes. *International Journal of Diabetes Mellitus*. 2010;3:189-95. doi: 10.1016/j.ijdm.2010.09.002.
15. Molteni R, Fabbri M, Bender JR, Pardi R. Pathophysiology of leukocyte-tissue interactions. *Curr Opin Cell Biol*. 2006 Oct;18(5):491-8. doi: 10.1016/j.ceb.2006.08.001.
16. Bertoni AG, Saydah S, Brancati FL. Diabetes and the risk of infection-related mortality in the U.S. *Diabetes Care*. 2001 Jun;24(6):1044-9. PMID: 11375368.

Received 25.06.2017 ■

Гавриленко Ю.В.

Національна медична академія післядипломної освіти імені П.Л. Шупика, м. Київ, Україна

### Вплив фітопрепарату BNO 1030 на фагоцитарну активність лейкоцитів крові при цукровому діабеті 1-го типу в щурів

**Резюме.** *Актуальність.* У патогенезі цукрового діабету 1-го типу (ЦД1) і багатьох його ускладнень важливе місце посідають порушення функціональної здатності імунної системи, що є предметом численних досліджень. З огляду на ефективність імуномодуючої терапії при ЦД1 і його ускладненнях однією з найважливіших проблем є пошук нових ефективних і безпечних імунокоригуючих засобів з високою фармакологічною активністю. **Мета дослідження:** оцінити вплив фітопрепарату BNO 1030 на фагоцитарну активність лейкоцитів крові при ЦД1 у щурів. **Матеріали та методи.** Експериментальний ЦД1 у щурів індукували шляхом одноразового внутрішньоочеревинного введення стрептозотину. Відсоток фагоцитуючих клітин і поглинання макрофагами живих флуоресцентних бактерій у зразках було визначено з використанням флуоресцентних живих бактерій *Escherichia coli*. **Результати.** На тлі гіперглікемії відбуваються зміни в перерозподілі основних типів лейкоцитів, а саме гранулоцитів і агранулоцитів. Введення препарату BNO 1030 як контрольним,

так і діабетичним тваринам приводило до збільшення кількості агранулоцитів, що може свідчити про модулюючий вплив препарату на імунну систему організму тварин. За цих умов знижувалася фагоцитарна активність лейкоцитів: фагоцитарне число в групі хворих на ЦД1 щурів менше ніж на 42 % було знижене порівняно з групою контрольних тварин. Уведення діабетичним щурам препарату BNO 1030 призводило до підвищення фагоцитарного числа на 24 % порівняно з групою діабетичних тварин. **Висновки.** Зміни супроводжувалися зниженням фагоцитарного індексу як при діабеті, так і при введенні препарату піддослідним групам, що свідчить про порушення в системі неспецифічного клітинного імунітету. Препарат BNO 1030 завдяки його імуномодуючій дії є ефективним засобом лікування цукрового діабету при його коморбідності з іншими хронічними захворюваннями.

**Ключові слова:** експериментальний цукровий діабет 1-го типу; BNO 1030; лейкоцити крові; фагоцитарна активність; фагоцитарне число; фагоцитарний індекс

Гавриленко Ю.В.

Национальная медицинская академия последипломного образования имени П.Л. Шупика, г. Киев, Украина

### Влияние фитопрепарата BNO 1030 на фагоцитарную активность лейкоцитов крови при сахарном диабете 1-го типа у крыс

**Резюме. Актуальность.** В патогенезе сахарного диабета 1-го типа (СД1) и многих его осложнений важное место занимают нарушения функциональной способности иммунной системы, что является предметом многочисленных исследований. С учетом эффективности иммуномодулирующей терапии при СД1 и его осложнениях одной из самых важных проблем является поиск новых эффективных и безвредных иммунокорректирующих средств с высокой фармакологической активностью. **Цель исследования:** оценить влияние фитопрепарата BNO 1030 на фагоцитарную активность лейкоцитов крови при СД1 у крыс. **Материалы и методы.** Экспериментальный СД1 у крыс индуцировали путем однократного внутрибрюшинного введения стрептозотоцина. Процент фагоцитирующих клеток и поглощения макрофагами живых флуоресцентных бактерий в образцах определяли с использованием флуоресцентных живых бактерий *Escherichia coli*. **Результаты.** На фоне гипергликемии происходят изменения в перераспределении основных типов лейкоцитов, а именно гранулоцитов и агранулоцитов. Введение препарата BNO 1030 как контрольным, так и диабетическим

животным приводило к увеличению количества агранулоцитов, что может свидетельствовать о модулирующем влиянии препарата на иммунную систему организма животных. В этих условиях снижалась фагоцитарная активность лейкоцитов: фагоцитарное число в группе больных СД1 крыс менее чем на 42 % было снижено по сравнению с группой контрольных животных. Введение диабетическим крысам препарата BNO 1030 приводило к повышению фагоцитарного числа на 24 % по сравнению с группой диабетических животных. **Выводы.** Изменения сопровождались снижением фагоцитарного индекса как при диабете, так и при введении препарата подопытным группам, что свидетельствует о нарушениях в системе неспецифического клеточного иммунитета. Препарат BNO 1030 благодаря его иммуномодулирующему действию является эффективным средством лечения сахарного диабета при его коморбидности с другими хроническими заболеваниями.

**Ключевые слова:** экспериментальный сахарный диабет 1-го типа; BNO 1030; лейкоциты крови; фагоцитарная активность; фагоцитарное число; фагоцитарный индекс